

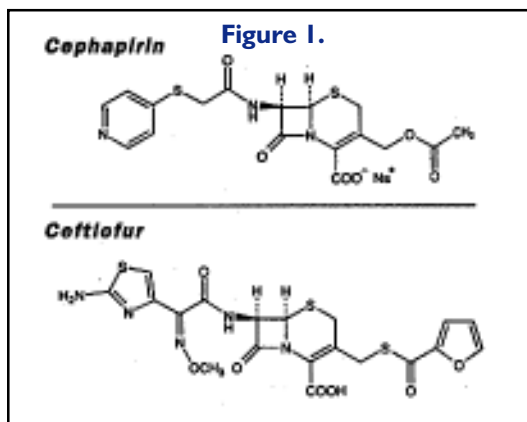
### Analysis of Antibacterial Residues in Cow Milk

Antibacteria are often used by farmers to treat diseases in dairy cow. While the treatment has been very effective in prevention of various diseases in cows and other dairy products, the effect of residues of these drugs in humans has recently become a topic of concern for the U.S. Food and Drug Administration (USFDA).

Ceftiofur and Cephapirin (Figure 1.) Are commonly used antibacterials. In vivo and in vitro antibacterial activity and B-lactamase stability of ceftiofur has been recently studied by Yancey et al.<sup>1</sup> Cephamycins represent a family of B-lactam antibiotics produced by various Streptomyces species. Cephapirin has been shown to be quite effective against gram-pos, and gram-neg, bacteria including Staphylococcus aureus, Escherichia coli, Klebsiella pneumoniae and Proteus mirabilis<sup>2</sup>. Because of the significant human consumption of dairy products, especially among infants, it is very crucial to monitor the concentration level of residues of these antibiotics in milk.

We report here a reversed-phase HPLC method that results in a very high resolution of Ceftiofur and Cephapirin in cow milk\*. Figure 2, shows the chromatogram of a: standard sample of 20 part per billion (ppb) of cephapirin and 50 ppb of ceftiofur compared with b: a chromatogram of extract from a donated raw sample from the State of South Dakota, USA. The antibacterials in milk were extracted using a number of liquid-liquid extractions followed by solid phase extraction and pre-concentration.

This method allows chromatographers to effectively isolate, characterize, and quantitate residues of the antibacterials in cow milk and possibly other dairy products.

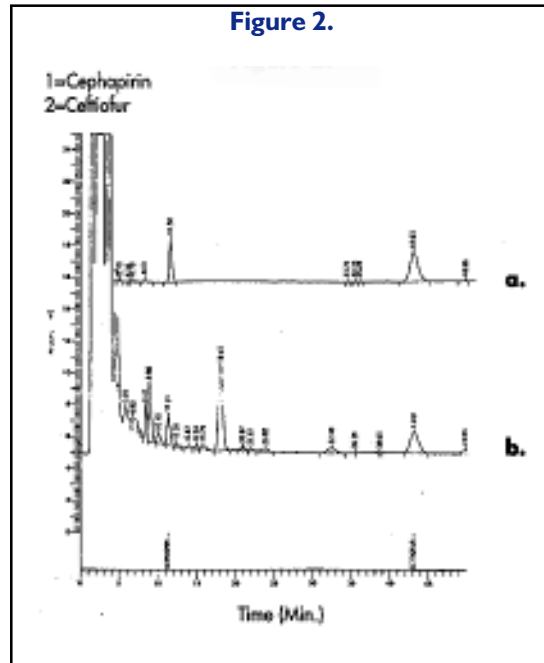


#### Chromatographic Conditions

Column: SMT 0-5-100  
Mobile Phase: 35 / 65 ACN / A  
A=9mM Sodiumdodecylsulfonic acid/18mM H<sub>3</sub>PO<sub>4</sub>  
Flow: 1 mL/min  
Inj: 200 µL  
Detector: UV, 290nm

#### Column Specifications:

Particle: Spherical silica, 5 µm  
Pore Size: 100 Å  
Surface Area: 340 m<sup>2</sup>/g  
Pore Volume: 1.0 ± 0.1  
% Carbon: 22%  
pH range: 1-12



\*SMT wishes to thank Dr. Pat Schermerhorn of U.S. FDA, Beltsville, MD for the method development.

1. Yancey et. al. Am. J. Vet. Res. 48, 1050, 1987.  
2. Crast et. al. J. Med. Chem. 16, 1413, 1973.

**SEPARATION  
METHODS  
TECHNOLOGIES**

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